

**Cytoplasmic c-myc Expression by Flow Cytometry**  
(Greg A. Perry, Ph.D.)

**Reagents:**

Perm-Wash buffer  
Fix-Perm reagent  
FACSFix

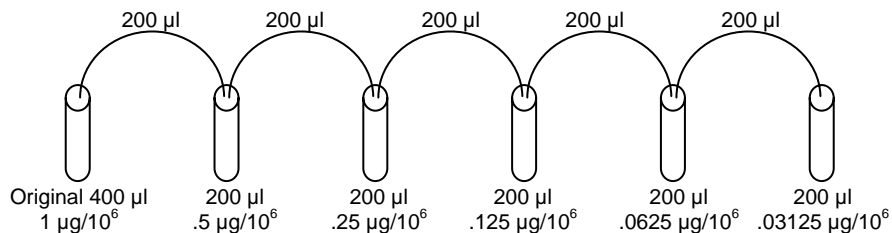
**Method:**

**I. Prepare Cells**

- Harvest cells (vector positive ( $v^+$ ) and vector negative ( $v^-$ ) 293 cells by gently washing off of dish.
- Centrifuge cells 10 minutes @ 350g (4°C).
- Remove supernatant and resuspend cells in 2ml PBS4.
- Adjust 293 cells to  $1 \times 10^6$  cells/ml in PBS4.
- Place 1 ml of the cells into 14 separate 12x75 mm tubes  
8 tubes for each cell type (labeled 1-8 for  $v^+$  and 9-16 for  $v^-$ )

**II. Prepare Antibody**

- Place 4  $\mu$ g (3.64  $\mu$ l) of antibody in an eppendorf tube.
  - Using anti-RAG #307
  - 1.1 mg/ml ( $\approx 1,100 \mu$ g/ml)
  - in 25mM Tris buffer (pH 7.6) with 150mM NaCl & 3mM KCl
- Add 20  $\mu$ l of the Zenon labeling reagent (Kit component A)
- Incubate at room temperature in the dark for 5 minutes.
- Add 20  $\mu$ l of the Zenon blocking reagent (Kit component B).
- Incubate at room temperature in the dark for 5 minutes.
- Bring up the total volume to 400  $\mu$ l by adding 376.4  $\mu$ l Perm/Wash buffer.
- Do doubling dilutions of the antibody in Perm/Wash buffer as follows:



**III. Fix, permeablize & stain the cells**

- Centrifuge cells 5 minutes @ **250g** (4°C).
- Remove supernatant.
- Remove tubes 1 ( $v^+$  cells only) and 9 ( $v^-$  cells only). Resuspend these two tubes in 1 ml FACSFix and place on ice.  
*All of the rest of the procedures will be performed on tubes 2-8 and 10-16 only.*
- Resuspend tubes 2-8 and 10-16 in 100  $\mu$ l of Fix&Perm reagent and mix gently.
- Incubate 20 minutes on ice.
- Add 1 ml Perm/Wash buffer and mix gently.
- Centrifuge 5 minutes @ 250g (4°C).
- Remove supernatant.

- Remove tubes 2 ( $v^+$  perm. cells only) and 10 ( $v^-$  perm. cells only). Resuspend these two tubes in 1 ml FACSFix and place on ice.

*All of the rest of the procedures will be performed on tubes 3-8 and 11-16 only.*

- Resuspend tubes 3-8 and 11-16 in 100  $\mu$ l of antibody as follows:
 

1 $\mu$ g/ $10^6$	tubes 3 and 11
.5 $\mu$ g/ $10^6$	tubes 4 and 12
.25 $\mu$ g/ $10^6$	tubes 5 and 13
.125 $\mu$ g/ $10^6$	tubes 6 and 14
.0625 $\mu$ g/ $10^6$	tubes 7 and 15
.03125 $\mu$ g/ $10^6$	tubes 8 and 16
- Mix gently and incubate 30 minutes at room temperature in the dark.
- Add 1 ml Perm/Wash buffer and mix gently.
- Centrifuge 5 minutes @ 250g (4°C).
- Remove supernatant.
- Resuspend in 1 ml Perm/Wash buffer and mix gently.
- Centrifuge 5 minutes @ 250g (4°C).
- Remove supernatant.
- Resuspend in 1 ml Perm/Wash buffer and mix gently.
- Centrifuge 5 minutes @ 250g (4°C).
- Remove supernatant.
- Resuspend cells in 1 ml FACSFix.
- Analyze on the cytometer.