

## MTT Mitogen Assay in Microtiter Plates

(Greg A. Perry, Ph.D.; after Mossman (1983))

## Equipment:

Flat Bottom Microtiter Plate (Costar #3596) Dual beam ELISA reader

## Reagents:

RF10-M Media (Sterile)

Cell Preparation (Steriile)

- at  $5 \times 10^6$  cells/ml in RF10-M media

Mitogens (pre-diluted)

- Concanavalin A (Con-A) (Type IV; Sigma #C-2010)
- Phytohemaglutinin-A (PHA) (Type M; Gibco #10576-015 or Sigma #L-8902)
- Lipopolysaccharide (LPS) (Serotype 055:B5; Sigma #L-2880)

MTT Stock (5 mg/ml in PBS; keep in dark at 4°C. Stable for several weeks.) Acid-Alcohol (0.04N HCl in 2-propanol)

## Method:

- 1. Add 100  $\mu$ I of cells to each appropriate well of the microtiter plate. (2x10<sup>5</sup> cells per well)
- 2. Add 100 µl of mitogen (or media) to each appropriate well containing cells.
- 3. Incubate at  $37^{\circ}$ C in a humidified atmosphere of 5% CO<sub>2</sub> in air for 72 hours.
- 4. Resuspend cells in each well, and transfer 25 μl of cells from each well into a corresponding well on a new plate containing 75 μl of fresh RF10-M in each well.
- 5. Add 10 µl of MTT stock to each well and mix by tapping gently on the side of the tray.
- 6. Incubate for 4 hours at 37°C for cleavage of MTT to occur (optimal time may vary).
- 7. Add 100 µl of acid-alcohol to each well and mix by pipetting up and down several times.
- 8. Within an hour, measure the absorbance on an ELISA plate reader with a test wavelength of 570 nm and a reference wavelength of 630 nm.
- 9. Express results as Optical Density (OD).

<u>Note:</u> Each new batch of mitogen must be tittered to determine optimal concentration for stimulation. Typical concentrations would be:

Con-A (Sigma, Type IV): ~ 5 μg/ml PHA (Wellcome): ~ 5 μg/ml LPS (Sigma, Serotype #055:B5): ~ 40 μg/ml

<u>Reference:</u> Mossman, T. (1983) Rapid calorimetric assay for cellular growth and survival: application to proliferation and cytotoxic assays. Journal of Immunological Methods 65: 55-63.